In-line Uranium Immunosensor

Diane A. Blake and Haini Yu, Tulane University Health Sciences Center, New Orleans, LA R. Mark Jones, Sapidyne Instruments Inc, Boise ID Robert C. Blake II, Xavier University of Louisiana, New Orleans, LA Immunosensors: Effective Analytical Tools for Environmental Analysis

- Provide rapid, near real-time results at the site of contamination
- Used to map contaminated sites and to quickly monitor the effectiveness of bioremediation and containment efforts
- Used in the analytical laboratory to rank samples by concentration before additional instrumental analysis

Antibody-Based Assays Are Cost-Effective

- Sample analysis is one of the major costs associated with the remediation of a contaminated site
- Studies have shown that the use of antibodybased assays can reduce analysis costs by 50% or more

Antibody-Based Assays Are Simple to Perform

- No complicated, sophisticated instruments are required
- Individuals performing the assays do not need to be highly skilled in analytical techniques
- Sample preparation procedures are usually uncomplicated and require minimal solvent

Project Goals

- To develop an in-line uranium immunosensor that can be used to determine the efficacy of specific biostimulation processes
- The sensor will be designed to autonomously:
- Self-calibrate using standard reagents;
- Collect a sample from a process line;
- Measure U(VI) at varying times during the treatment process.



Optics, LED, and capillary bed containing particles with immobilized capture ligand.

 Fluorescently labeled antibody and environmental contaminant are mixed in a disposable syringe.

Drive motor pushes the antibody-contaminant mixture over the capture ligand. Generation of UO_2^{2+} -specific monoclonal antibodies using a 2,9-dicarboxyl-1,10-phenanthroline (DCP) derivative



- 1. Prepare DCP-protein conjugate
- 2. Load with UO_2^{2+}
- 3. Inject mice and wait for immune response to develop
- 4. Harvest spleen cells and make hybridomas
- 5. Screen hybridomas for antibodies with desirable characteristics for incorporation into sensors

Desirable Characteristics for Metal-Specific Antibodies

- Little or no binding to the metal-free chelator
- Tight binding to the chelated metal of interest
- Little or no binding to other metals most likely to contaminate the environmental sample
- Little or no interference from components likely to be present in the environmental sample matrix

Equilibrium dissociation constants for the binding of 2,9dicarboxyl-1,10-phenanthroline (DCP) in the presence and absence of uranyl ion to monoclonal antibodies 12F6, 10A3, and 8A11.

value of K _d , M			
Ligand	12F6	10A3	8A11
N — UO ₂ ²⁺ СО ₂ Н	9.1 ± 0.7 × 10 ⁻¹⁰	2.4 ± 0.2 × 10 ⁻⁹	5.5 ± 0.2 × 10 ⁻⁹
Metal-free DCP	7.5 ± 0.5 × 10 ⁻⁷	2.8 ± 0.1 × 10 ⁻⁶	3.7 ± 0.2 × 10 ⁻⁶



Keys to Successful Immunosensor Development

- Inclusion of the antibodies and the sample matrix to be used in the final assay early in sensor development.
- Exhaustive characterization of the binding properties of the antibody being used in the sensor.

Other factors influencing the performance of the uranium immunosensor

- BSA, a carrier protein added to the assay system to improve antibody stability, bound the UO₂²⁺-DCP complex with micromolar affinity (K_d of 3.3 ×10⁻⁶ M) and caused the assay to appear less sensitive for uranium.
- Covalent modification of the lysine residues of 12F6 destroyed this antibody's binding activity.
- Covalent modification of the lysine residues of 8A11 with Cy5 or Alexa 488 induced positive cooperativity in its binding to the UO₂²⁺-DCP complex (Hill coefficient of 1.5-1.6).
- Incubation of 8A11 with saturating concentrations of fluorescently-labeled goat antibodies directed against mouse IgG increased the affinity of the native 8A11 for the UO₂²⁺-DCP complex by three-fold.



Detailed binding studies performed for immunosensor development have revealed basic information about immunoglobulin structure and function heretofore unrecognized in the field.





Table 1. Analytical Recovery of U(VI) added to water samples

found U(VI), nM	recovery (%)
3.107 <u>+</u> 0.203	77.67 <u>+</u> 6.53
7.032 <u>+</u> 0.241	93.75 <u>+</u> 3.43
13.008 <u>+</u> 0.339	104.06 <u>+</u> 2.60
15.821 <u>+</u> 1.881	105.48 <u>+</u> 11.9
19.064 <u>+</u> 2.136	105.91 <u>+</u> 11.2
21.634 <u>+</u> 1.435	108.17 <u>+</u> 6.63
	99.17 <u>+</u> 7.05
	found U(VI), nM 3.107 <u>+</u> 0.203 7.032 <u>+</u> 0.241 13.008 <u>+</u> 0.339 15.821 <u>+</u> 1.881 19.064 <u>+</u> 2.136 21.634 <u>+</u> 1.435

Autonomous dilution and subsequent assay of an NIST uranium standard by the in-line immunosensor



Manual dilution and subsequent assay of an NIST uranium standard with KPA





Cloning Antibody 12F6



Cloning Antibody 12F6



Dicistronic vector used for recombinant antibody expression



Binding of culture supernatants from transfected cells to immobilized UO_2^{2+} -DCP



Conclusions

- Previous studies on antibody characterization and instrument development streamlined the development of an in-line immunosensor for hexavalent uranium.
- The LOD of the assay is 5.8 nM (1.38 ppb), with CV's from 2.3-13.9%.
- Expression of the 12F6 antibody as a recombinant protein will facilitate its incorporation into the sensor format.

Future Studies

- Deploy the in-line sensor at other places in the DOE complex
- Develop new assays for additional heavy metals, Hg(II) and Cr(III)
- Develop new recombinant antibody reagents based on antibody phage display techniques



Acknowledgements

Craig Criddle, Stanford University

Dave Watson and Scott Brooks, ORNL

Steve Lackie, Sapidyne Instruments, Boise ID

US-EC Biotechnology Fellowship Program (Judy Wall and Joe Suflita)

www.som.tulane.edu/labs/blake